

ap biology lab eight population genetics evolution answers

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Mastering AP Biology Lab 8: Population Genetics and Evolution – Complete Answers and Insights

AP Biology Lab 8 is a cornerstone of the Advanced Placement curriculum, bridging the gap between textbook theory and hands on scientific inquiry. This laboratory focuses on population genetics and evolution, challenging students to collect data, calculate allele frequencies, and interpret the forces that shape genetic diversity. Whether you're a teacher preparing a unit, a student tackling the exam, or an educator looking for fresh teaching ideas, this guide provides a comprehensive, step by step walkthrough of the lab's objectives, methodology, data analysis, and key conceptual take aways. Below, you'll find a detailed explanation of every component, along with sample answers to the typical questions that appear on the AP exam.

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1. Engaging Introduction

Imagine a population of organisms that, over successive generations, changes in size, structure, and genetic makeup. In the natural world, these shifts are driven by forces such as natural selection, genetic drift, mutation, and gene flow. AP Biology Lab 8 invites you to witness these forces in action through a controlled experiment that mirrors evolutionary dynamics. By measuring allele frequencies across generations, students gain a tangible understanding of how populations evolve—an experience that not only solidifies core genetics concepts but also hones essential scientific skills: data collection, statistical reasoning, and critical interpretation.

2. Lab Objectives

The primary goals of Lab 8 are to:

1. Determine allele and genotype frequencies in a population.
2. Assess whether the population adheres to Hardy Weinberg equilibrium.
3. Explore the impact of genetic drift and selection on allele frequencies.
4. Develop proficiency in data analysis, including chi square tests and graphical representation.
5. Apply evolutionary principles to real world scenarios.

3. Materials and Setup

While the actual laboratory can vary based on institutional resources, the most common setup involves:

- Organisms with easily scorable phenotypes (e.g., *Caenorhabditis elegans*, *E. coli*, or *Arabidopsis thaliana*)
- Growth media (agar plates, nutrient broth, etc.)
- Microscopes or stereomicroscopes for phenotype observation
- Stirring rods, pipettes, and sterile gloves
- Data sheets or digital spreadsheets for recording counts
- Statistical software or graphing calculators for chi square and frequency calculations

4. Experimental Procedure

The lab typically unfolds over several days, allowing the population to undergo multiple generations. Below is a generalized protocol:

1. Initial Population Setup: Prepare a starting population with known genotype frequencies. For example, if studying a single gene with two alleles (A and a), seed the population with a mixture such that you can calculate initial frequencies.
2. Cultivation: Grow organisms under controlled conditions (temperature, light, nutrients). Ensure that the environment remains constant to isolate genetic factors.
3. Generational Transfer: At the end of each generation, transfer a fixed number of individuals to fresh media. This step simulates population bottlenecks and can influence genetic drift.
4. Phenotype Scoring: Observe and record the phenotype of each individual. Convert phenotypes to genotypes using Mendelian ratios (e.g., dominant vs. recessive).
5. Data Recording: Tally counts of each genotype and calculate allele frequencies (p and q) for each generation.
6. Repeat: Continue this process for at least three generations to observe trends.

5. Data Collection & Analysis

Accurate data analysis is the linchpin of this lab. The steps below guide you from raw counts to meaningful evolutionary insights.

5.1 Calculating Allele Frequencies

For a gene with two alleles (A and a), the allele frequencies are calculated as:

$$p = (2 * \#AA + \#Aa) / (2 * N)$$

$$q = (2 * \#aa + \#Aa) / (2 * N)$$

where **N** is the total number of individuals. Verify that **p + q = 1** within rounding error.

5.2 Hardy Weinberg Equilibrium (HWE)

Under HWE, genotype frequencies are predicted by:

$$f(AA) = p^2$$

$$f(Aa) = 2pq$$

$$f(aa) = q^2$$

Use a chi square test to compare observed vs. expected genotype counts:

$$\chi^2 = \sum [(O - E)^2 / E]$$

With one degree of freedom (since allele frequencies are estimated from data), a p value greater than 3.84 (p

5.3 Detecting Genetic Drift

Genetic drift is most pronounced in small populations. To assess drift:

- Plot allele frequency changes across generations.
- Calculate the standard deviation of allele frequencies.
- Compare observed variance to the theoretical variance predicted by the binomial model: $\text{Var}(p) = pq / (2N)$.

5.4 Evaluating Selection

Selection can be inferred when genotype frequencies deviate from HWE in a systematic way. For example, if the homozygous dominant genotype consistently appears at a higher frequency than expected, this suggests positive selection for that allele.

5.5 Graphical Representation

Visualizing data helps in interpretation:

- Allele Frequency Bar Graphs – Show p and q over generations.
- Genotype Frequency Pie Charts – Illustrate the proportional makeup of the population.
- Chi Square Line Graphs – Track the significance of deviations over time.

6. Key Genetic Concepts Covered

- Allele vs. Genotype – Understanding the difference between the gene variants and the organism's genetic constitution.
- Hardy Weinberg Principle – The baseline expectation for allele and genotype frequencies in an ideal population.
- Genetic Drift – Random fluctuations in allele frequencies, especially in small populations.
- Selection – Non random survival or reproduction that favors certain genotypes.
- Mutation – Introduction of new alleles, albeit at low rates in most lab settings.
- Gene Flow – Migration of alleles between populations, not typically modeled in Lab 8 but essential for broader evolutionary understanding.

7. Sample Answers to Common Lab 8 Questions

Below are concise, well structured responses to typical AP Biology Lab 8 exam questions. These answers illustrate how to integrate data with theory.

7.1 Question: “Describe the changes in allele frequencies over the three generations. What evolutionary forces might explain these changes?”

Answer: The allele frequency of A increased from 0.55 to 0.68 over three generations, while a decreased from 0.45 to 0.32. This upward trend in p suggests a selective advantage for the dominant allele, potentially due to higher fitness in the given environment. Alternatively, genetic drift could account for the shift if the population size was small; however, the consistent increase across all replicates points more strongly toward directional selection.

7.2 Question: “Perform a chi square test to determine if the population is in Hardy Weinberg equilibrium during Generation 2. Provide the expected genotype counts, chi square value, and interpretation.”

Answer: In Generation 2, observed counts were: AA = 30, Aa = 40, aa = 30 ($N = 100$). Calculated allele frequencies: $p = 0.55$, $q = 0.45$. Expected genotype counts: $f(\text{AA}) = p^2 = 0.3025 \rightarrow 30.25$, $f(\text{Aa}) = 2pq = 0.495 \rightarrow 49.5$,

$f(aa) = q^2 = 0.2025$!' 20.25. Chi square: $\chi^2 = (30-30.25)^2/30.25 + (40-49.5)^2/49.5 + (30-20.25)^2/20.25 = 0.003 + 1.728 + 4.354 = 6.085$. With one degree of freedom, $\chi^2 = 6.085 > 3.84$, indicating a significant deviation from HWE. This suggests selection or non random mating is influencing genotype frequencies.

7.3 Question: “Explain how genetic drift can be distinguished from selection in this experiment.”

Answer: Genetic drift manifests as random, unpredictable changes in allele frequencies, often leading to fixation or loss of alleles in small populations. In contrast, selection produces consistent, directional changes that correlate with fitness differences. In Lab 8, if allele frequency changes are similar across all replicates and correlate with a measurable fitness advantage (e.g., faster growth), selection is the likely cause. Random fluctuations that vary widely between replicates, especially in small populations, point to drift.

7.4 Question: “Calculate the effective population size (Ne) given the observed variance in allele frequency between two generations.”

Answer: Observed variance (σ^2) in allele frequency between Generation 1 ($p = 0.50$) and Generation 2 ($p = 0.55$) was 0.025. Using the formula $\sigma^2 = pq/(2N_e)$, we solve for $N_e = pq/(2\sigma^2) = (0.50 \cdot 0.50)/(2 \cdot 0.025) = 0.25/0.05 = 5$. Thus, the effective population size is approximately 5 individuals, indicating strong drift potential.

7.5 Question: “Propose a modification to the experimental design that would allow you to isolate the effect of mutation.”

Answer: To isolate mutation, increase the number of generations and maintain a large population size to minimize drift. Introduce a mutagenic agent (e.g., UV light or chemical mutagens) at a controlled dose and compare the mutant frequency to a control group without mutagen exposure. By measuring new allele appearance rates and ensuring other variables remain constant, the impact

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